

## Appendix No. 1 – ML 4912/21

Records documenting analysis of the sample using U-HPLC-HRMS/MS metabolomic profiling (targeted screening) method

### Sample description

Lab code	Sample name (provided by the client)	Sample description
ML 4912/21	Olejek konopny 10% CBG Dekoktum	Hemp oil

### Testing strategy

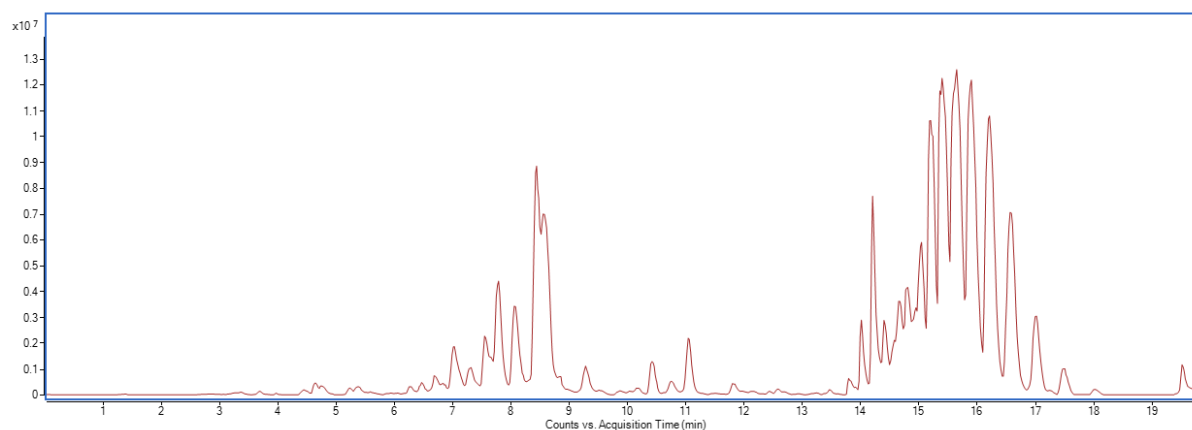
Metabolomic profiling (targeted screening) strategy was selected for the testing of the sample (laboratory code ML 4912/21). This type of analytical approach is based on UCT internal databases of compounds which may occur in *Cannabis sativa* L. and products thereof; the first UCT database (A) includes molecular spectral information for 246 minor phytocannabinoids, the second UCT database (B) includes molecular spectral information for 151 other (non-cannabinoid) biologically active compounds (e.g. terpenoids, phenols, bibenzyl stilbenes, fatty acids, amides, flavones, lignans, flavonoid glycosides, lignanamide derivatives). The analysis of the given sample was performed by ultra-high performance liquid chromatography coupled to high resolution tandem mass spectrometry (ISO 17025 accredited method KM 15, system E: U-HPLC-HRMS/MS (Q-TOF)). Detailed description of analytical procedures and conditions (SOP) are available at Laboratory.

### Testing conditions

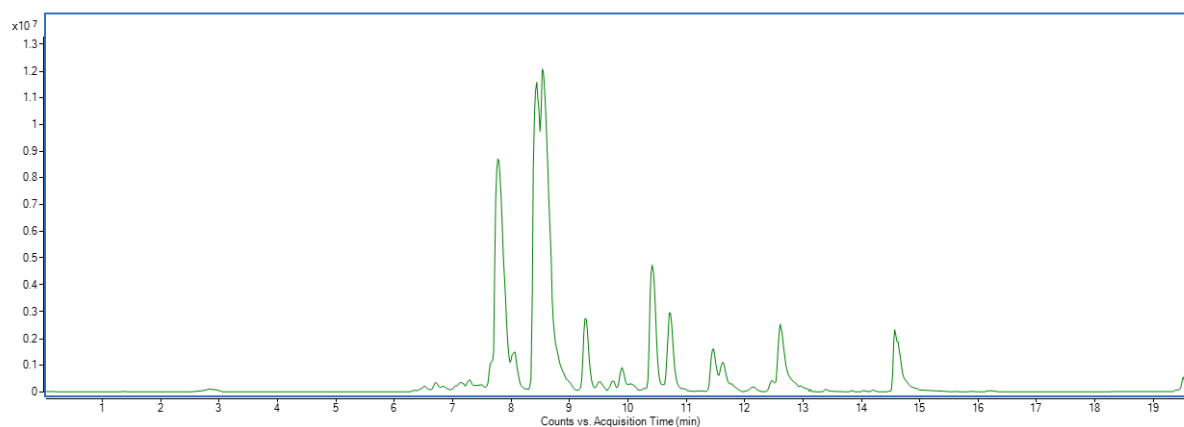
Sample preparation consisted of sample dissolution in ethanol and following dilution. Subsequently, a reversed-phase chromatographic column was used for the compound's separation; quadrupole/time of flight mass analyzer (Agilent 6560 Q-TOF) with electrospray ionization source was used for their detection. Each sample was injected into U-HPLC – HRMS/MS system in several dilutions to overcome matrix effects including column and/or detector saturation for particular compounds. The data were recorded and evaluated separately for both the positive and negative electrospray ionization mode (ESI+ and ESI-). For the data processing, Agilent MassHunter Profinder and Agilent MassHunter Qualitative Analysis softwares were used.

### Test results and interpretation

The overall chemical compositions of the sample detectable under given method conditions are illustrated by total compound chromatograms (TCC) (**Figure 1 - 2**) recorded in ESI+ and ESI- ionization mode. The TCC shows all features (compounds) detected in the sample, non-filtered by the UCT databases.



**Figure 1:** ML 4912/21 - UHPLC-HRMS/MS total compound chromatogram obtained for undiluted sample in ESI+



**Figure 2:** ML 4912/21 - UHPLC-HRMS/MS total compound chromatogram obtained for undiluted sample in ESI-

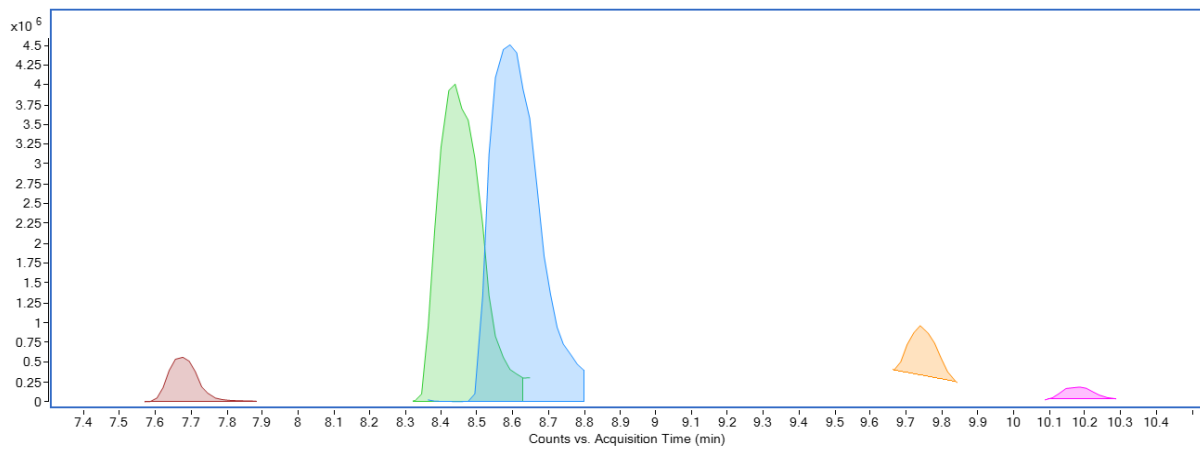
#### A) Targeted screening of phytocannabinoids

The following amounts of major phytocannabinoids (**Table I**) characterized by unique combination of exact mass and retention time RT, whose identities were confirmed by certified standards, were detected.

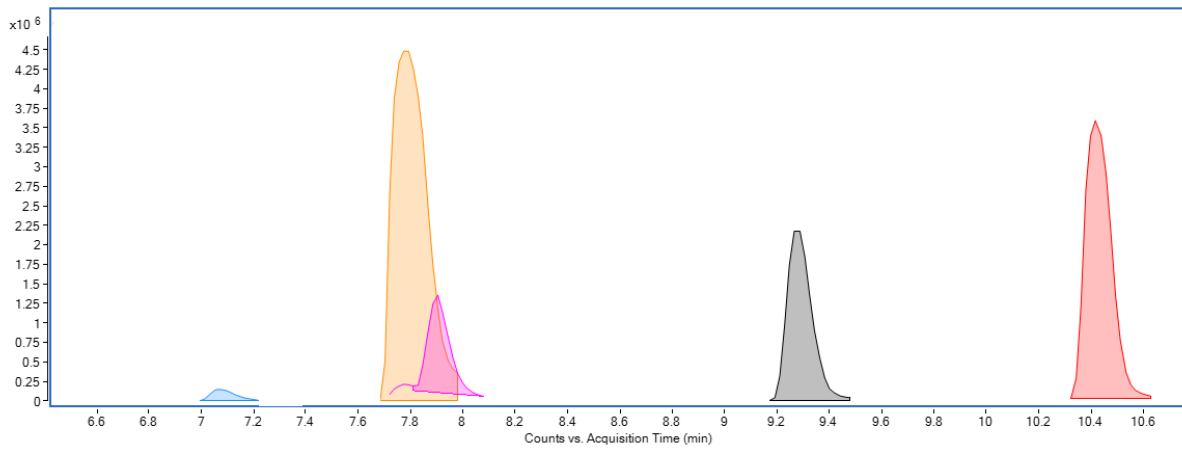
**Table I:** Number of major phytocannabinoids detected in the sample in ESI+ and ESI-

Sample name	Number of major phytocannabinoids detected in ESI+	Number of major phytocannabinoids detected in ESI-
ML 4912/21	5	5

Extracted ion chromatograms (XIC) of these compounds in both ionization modes (ESI+, ESI-) are documented on **Figures 3 – 4** and their overview is also summarized in **Table II**.



**Figure 3:** ML 4912/21 - UHPLC-HRMS/MS extracted ion chromatogram of major phytocannabinoids detected in ESI+ (obtained for undiluted sample)



**Figure 4:** ML 4912/21 - UHPLC-HRMS/MS extracted ion chromatogram of major phytocannabinoids detected in ESI- (obtained for undiluted sample)

**Table II:** Overview of major phytocannabinoids detected in ESI+ and ESI- (corresponding with the Figures 3 - 4)

Measured exact mass <sup>a</sup> (neutral, monoisotopical)	Elemental formula <sup>b</sup>	RT <sup>c</sup> (min)	Ionization mode	Identity (confirmed by certified standards)	Peak area <sup>d</sup>
330.1832	C20 H26 O4	7.08	ESI-	CBDVA	1.0E+06
286.1937	C19 H26 O2	7.67	ESI+	CBDV	3.3E+06
358.2143	C22 H30 O4	7.79	ESI-	CBDA	6.2E+07
360.2296	C22 H32 O4	7.91	ESI-	CBGA	7.4E+06
316.2398	C21 H32 O2	8.44	ESI+	CBG	1.3E+08
314.2252	C21 H30 O2	8.58	ESI+	CBD	1.8E+08
310.1933	C21 H26 O2	9.28	ESI-	CBN	1.5E+07
314.2253	C21 H30 O2	9.74	ESI+	$\Delta^9$ -THC	5.2E+06
314.2251	C21 H30 O2	10.18	ESI+	CBL	1.0E+06
314.2248	C21 H30 O2	10.43	ESI-	CBC	2.6E+07

<sup>a</sup> Mass error compared the theoretical exact mass < 5 ppm

<sup>b</sup> Match of isotopic pattern confirmed

<sup>c</sup> Retention time

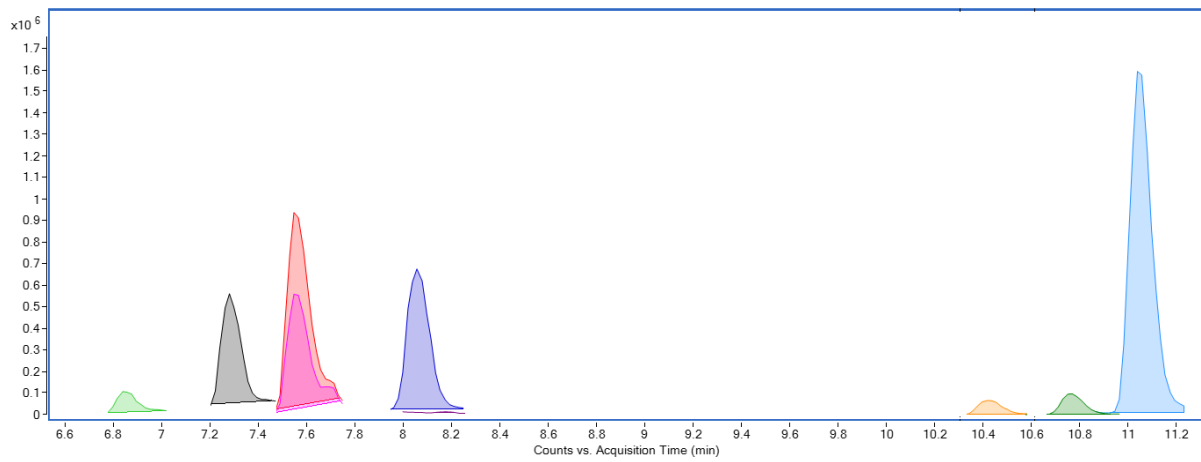
<sup>d</sup> Peak area related to the undiluted sample prepared according to stated procedure

The following amounts of minor phytocannabinoids (**Table III**) characterized by unique combination of exact mass  $m/z$  and retention time RT were detected.

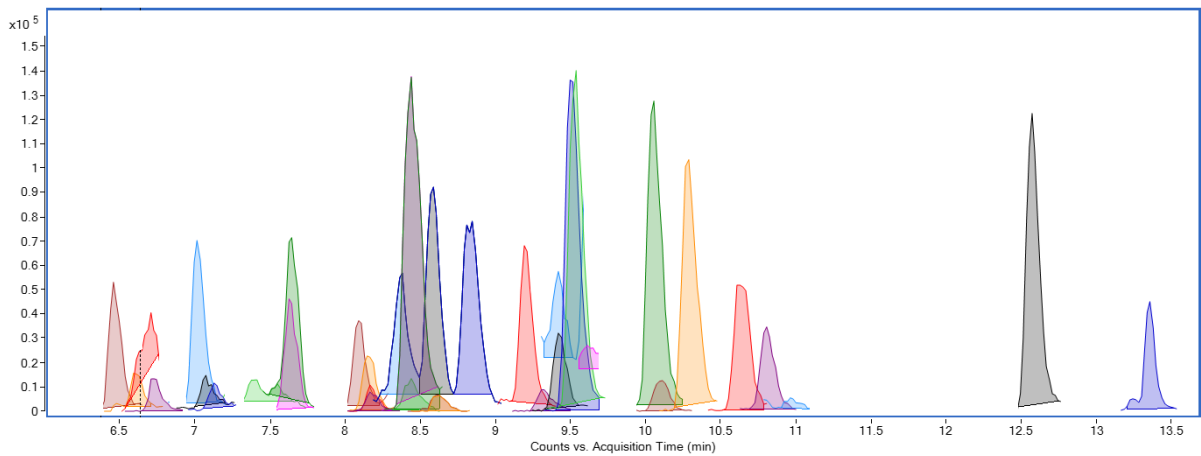
**Table III:** Number of minor phytocannabinoids detected in the sample in ESI+ and ESI-

Sample name	Number of minor phytocannabinoids detected in ESI+	Number of minor phytocannabinoids detected in ESI-
ML 4912/21	9	43

Extracted ion chromatograms (XIC) of the detected compounds in both ionization modes (ESI+, ESI-) are documented on **Figures 5 - 6**. All detected phytocannabinoids (tentative identification) are also summarized in **Table IV** (ESI+) and **Table V** (ESI-). Considering the possible existence of isomeric forms, one detected compound may have several identities.



**Figure 5:** ML 4912/21 - UHPLC-HRMS/MS extracted ion chromatogram of 9 compounds with unique combination of exact mass and RT detected in ESI+ (obtained for undiluted sample)



**Figure 6:** ML 4912/21 - UHPLC-HRMS/MS extracted ion chromatogram of 43 compounds with unique combination of exact mass and RT detected in ESI- (obtained for undiluted sample)

**Table IV:** Overview of compounds detected in ESI+ (corresponding with the Figure 5)

Cmp. number	Measured exact mass <sup>a</sup> (neutral, monoisotopical)	Estimated elemental formula <sup>b</sup>	RT <sup>c</sup> (min)	Tentative identity (name of possible compounds) <sup>d</sup>	Peak area <sup>e</sup>
1	332.2350	C <sub>21</sub> H <sub>32</sub> O <sub>3</sub>	6.84	rac-6'-Epoxy <span>cannabigerol</span> (2'S*. 3'R*)/rac-6'-epoxy <span>cannabigerol</span> (2'R*. 3'R*)/(-)-7-Hydroxy <span>cannabichromane</span>	5.9E+05
2	330.2200	C <sub>21</sub> H <sub>30</sub> O <sub>3</sub>	7.28	8'-Hydroxy <span>isocannabichromene</span> /cannabielsoin/abnormal <span>cannabigerol</span> quinol/10alpha-hydroxy trans delta-8-tetrahydro <span>cannabinol</span> /10 beta-hydroxy trans delta-8-tetrahydro <span>cannabinol</span> /8 alpha-hydroxy-delta9-trans-tetrahydro <span>cannabinol</span> /8 beta-hydroxy-delta9-trans-tetrahydro <span>cannabinol</span> /tetrahydro <span>cannabinol</span> epoxide/hydroxy delta9.11-hexahydro <span>cannabinol</span>	2.9E+06
3	314.2254	C <sub>21</sub> H <sub>30</sub> O <sub>2</sub>	7.55	<span>Cannabicitran</span> /(-)-delta9-cis-(6aS.10aR)-delta9-tetrahydro <span>cannabinol</span> /(\hat{a}')-delta7 -trans-(1R. 3R. 6R)-isotetrahydro <span>cannabinol</span> -C5/ <span>cannabigerol</span> quinone	7.1E+06
4	332.2357	C <sub>21</sub> H <sub>32</sub> O <sub>3</sub>	7.55	rac-6'-Epoxy <span>cannabigerol</span> (2'S*. 3'R*)/rac-6'-epoxy <span>cannabigerol</span> (2'R*. 3'R*)/(-)-7-Hydroxy <span>cannabichromane</span>	4.3E+06
5	330.2203	C <sub>21</sub> H <sub>30</sub> O <sub>3</sub>	8.06	8'-Hydroxy <span>isocannabichromene</span> /cannabielsoin/abnormal <span>cannabigerol</span> quinol/10alpha-hydroxy trans delta-8-tetrahydro <span>cannabinol</span> /10 beta-hydroxy trans delta-8-tetrahydro <span>cannabinol</span> /8 alpha-hydroxy-delta9-trans-tetrahydro <span>cannabinol</span> /8 beta-hydroxy-delta9-trans-tetrahydro <span>cannabinol</span> /tetrahydro <span>cannabinol</span> epoxide/hydroxy delta9.11-hexahydro <span>cannabinol</span>	4.3E+06
6	314.2247	C <sub>21</sub> H <sub>30</sub> O <sub>2</sub>	8.18	<span>Cannabicitran</span> /(-)-delta9-cis-(6aS.10aR)-delta9-tetrahydro <span>cannabinol</span> /(\hat{a}')-delta7 -trans-(1R. 3R. 6R)-isotetrahydro <span>cannabinol</span> -C5/ <span>cannabigerol</span> quinone	4.4E+04
7	258.1622	C <sub>17</sub> H <sub>22</sub> O <sub>2</sub>	10.42	<span>Cannabidiol</span> col/delta-9-trans-tetrahydro <span>cannabiorcol</span> / <span>cannabiorcitrin</span> / <span>cannabiorcyclo</span> l/ <span>cannabiorcichromene</span>	4.8E+05
8	330.2559	C <sub>22</sub> H <sub>34</sub> O <sub>2</sub>	10.77	O-Methyl <span>cannabigerol</span>	6.4E+05
9	314.2252	C <sub>21</sub> H <sub>30</sub> O <sub>2</sub>	11.05	<span>Cannabicitran</span> /(-)-delta9-cis-(6aS.10aR)-delta9-tetrahydro <span>cannabinol</span> /(\hat{a}')-delta7 -trans-(1R. 3R. 6R)-isotetrahydro <span>cannabinol</span> -C5/ <span>cannabigerol</span> quinone	1.0E+07

<sup>a</sup> Mass error compared the theoretical exact mass < 5 ppm

<sup>b</sup> Match of isotopic pattern confirmed

<sup>c</sup> Retention time

<sup>d</sup> Tentative identification of compounds based on available scientific articles (Hanusš et al. 2016, Mechoulam 2002); possible identities are separated by the symbol "/"

<sup>e</sup> Peak area related to the undiluted sample prepared according to stated procedure

**Table V:** Overview of compounds detected in ESI- (corresponding with the Figure 6)

Cmp. number	Measured exact mass <sup>a</sup> (neutral, monoisotopical)	Estimated elemental formula <sup>b</sup>	RT <sup>c</sup> (min)	Tentative identity (name of possible compounds) <sup>d</sup>	Peak area <sup>e</sup>
1	330.1822	C20 H26 O4	5.69	Cannabichromevarinic acid/delta-9-tetrahydrocannabivarinic acid	9.4E+03
2	282.1617	C19 H22 O2	6.46	Cannabinodivarin/cannabivarin/demethyldecarb oxyamorfrutin A/63b/63c	3.1E+05
3	376.2241	C22 H32 O5	6.59	rac-6'-Epoxyacnabigerolic acid (2'S*. 3'R*)/rac-6'-epoxyacnabigerolic acid (2'R*. 3'R*)	8.9E+04
4	332.2344	C21 H32 O3	6.71	rac-6'-Epoxyacnabigerol (2'S*. 3'R*)/rac-6'-epoxyacnabigerol (2'R*. 3'R*)/(-)-7-Hydroxyacnabichromane	3.3E+05
5	294.1616	C20 H22 O2	6.73	Radulanin J	8.4E+04
6	374.2092	C22 H30 O5	7.02	Cannabielsoic acid A/cannabielsoic acid B	4.0E+05
7	302.1870	C19 H26 O3	7.07	C3-Cannabielsoin	7.8E+04
8	358.2138	C22 H30 O4	7.14	Ferruginene A/ferruginene B/delta-9-tetrahydrocannabinolic acid B/delta-8-tetrahydrocannabinolic acid	7.0E+04
9	348.2290	C21 H32 O4	7.38	Cannabiripsol	5.4E+04
10	348.2290	C21 H32 O4	7.55	Cannabiripsol	6.6E+04
11	310.1929	C21 H26 O2	7.64	Cannabinodiol/cannabifuran	2.8E+05
12	288.2082	C19 H28 O2	7.65	Cannabigerovarin	4.4E+05
13	300.2088	C20 H28 O2	7.90	delta-9-Tetrahydrocannabinol-C4/norCannabidiol	7.0E+06
14	300.2083	C20 H28 O2	8.10	delta-9-Tetrahydrocannabinol-C4/norCannabidiol	7.0E+05
15	316.2394	C21 H32 O2	8.10	Cannabinerol/abnormal cannabigerol/hexahydrocannabinol	2.1E+05
16	282.1618	C19 H22 O2	8.15	Cannabinodivarin/cannabivarin/demethyldecarb oxyamorfrutin A/63b/63c	1.4E+05
17	366.2185	C24 H30 O3	8.16	Hydroxy helicannabigenol/55	6.0E+04
18	258.1617	C17 H22 O2	8.17	Cannabidiolcol/delta-9-trans-tetrahydrocannabinolcol/cannabiorcitrin/cannabiorcicyclol/cannabiorcichromene	4.5E+04
19	332.2345	C21 H32 O3	8.37	rac-6'-Epoxyacnabigerol (2'S*. 3'R*)/rac-6'-epoxyacnabigerol (2'R*. 3'R*)/(-)-7-Hydroxyacnabichromane	3.6E+05
20	308.1774	C21 H24 O2	8.44	Dehydrocannabifuran	9.7E+04
21	348.2288	C21 H32 O4	8.44	Cannabiripsol	1.1E+06
22	288.2081	C19 H28 O2	8.45	Cannabigerovarin	2.6E+06
23	332.2346	C21 H32 O3	8.59	rac-6'-Epoxyacnabigerol (2'S*. 3'R*)/rac-6'-epoxyacnabigerol (2'R*. 3'R*)/(-)-7-Hydroxyacnabichromane	5.5E+05
24	318.1825	C19 H26 O4	8.61	(9S.10S)-trans-Cannabitrinol-C3/(9R.10R)-trans-cannabitrinol-C3	5.2E+04
25	258.1620	C17 H22 O2	8.63	Cannabidiolcol/delta-9-trans-tetrahydrocannabinolcol/cannabiorcitrin/cannabiorcicyclol/cannabiorcichromene	4.6E+04

**Table V - continuation:** Overview of compounds detected in ESI- (corresponding with the Figure 6)

Cmp. number	Measured exact mass <sup>a</sup> (neutral, monoisotopical)	Estimated elemental formula <sup>b</sup>	RT <sup>c</sup> (min)	Tentative identity (name of possible compounds) <sup>d</sup>	Peak area <sup>e</sup>
26	332.2347	C21 H32 O3	8.84	rac-6'-Epoxy cannabigerol (2'S*. 3'R*)/rac-6'-epoxy cannabigerol (2'R*. 3'R*)/(-)-7-Hydroxycannabichromane	6.2E+05
27	316.2401	C21 H32 O2	8.85	Cannabinol/abnormal cannabigerol/hexahydrocannabinol	3.6E+06
28	286.1929	C19 H26 O2	9.20	(delta)-7-trans-(1R, 3R, 6R)-isotetrahydrocannabivarin-C3 (trans-Isotetrahydrocannabivarin)/(1aS,3aR,8bR,8cR)-Cannabicyclovarin/Cannabivarinchromene ((RS)-Cannabichromevarin)/delta-9-trans-tetrahydrocannabivarin/2-Methyl-2-(4-methyl-2-pentenyl)-7-propyl-2H-1-benzopyran-5-ol/delta-7-1.2-cis-(1R,3R,6S)-Isotetrahydrocannabivarin-C3 (7-cis-Isotetrahydrocannabivarin)/delta-7-1.2-cis-(1S,3S,6R)-Isotetrahydrocannabivarin-C3/delta-9-cis-Tetrahydrocannabidivarin	3.7E+05
29	316.2390	C21 H32 O2	9.30	Cannabinol/abnormal cannabigerol/hexahydrocannabinol	4.9E+05
30	268.1467	C18 H20 O2	9.31	Cannabinol-C2/56a/63a	6.9E+04
31	314.2241	C21 H30 O2	9.43	Cannabicitran/(-)-delta-9-cis-(6aS,10aR)-delta-9-tetrahydrocannabinol/(â)-delta-7-trans-(1R, 3R, 6R)-isotetrahydrocannabinol-C5/cannabigeronequinone	2.2E+05
32	374.2451	C23 H34 O4	9.43	(-)-(9R,10R)-trans-10-O-Ethylcannabitol/5-acetyl-4-hydroxycannabigerol/acetyl abnormal cannabigeronequinol/cannabigeronic acid monomethylether	2.1E+05
33	342.2190	C22 H30 O3	9.51	Ferruginene C/2-formyl-delta-9-trans-tetrahydrocannabinol	9.4E+05
34	328.2037	C21 H28 O3	9.53	Cannabichromanone-D/cannabicomaronone/10-oxo-delta-6a(10a)-tetrahydrocannabinol/8-Oxo-delta-9-trans-tetrahydrocannabinol/9.10-Anhydrocannabitol/anhydrocannabimovone/Cannabidiol Hydroxyquinone	9.3E+05
35	316.2389	C21 H32 O2	9.75	Cannabinol/abnormal cannabigerol/hexahydrocannabinol	9.8E+04
36	330.2192	C21 H30 O3	10.06	8'-Hydroxyisocannabichromene/cannabielsoin/abnormal cannabigeronequinol/10alpha-hydroxy trans delta-8-tetrahydrocannabinol/10 beta-hydroxy trans delta-8-tetrahydrocannabinol/8 alpha-hydroxy-delta-9-trans-tetrahydrocannabinol/8 beta-hydroxy-delta-9-trans-tetrahydrocannabinol/tetrahydrocannabinol epoxide/hydroxy delta-9.11-hexahydrocannabinol	8.5E+05



**Table V - continuation:** Overview of compounds detected in ESI- (corresponding with the Figure 6)

Cmp. number	Measured exact mass <sup>a</sup> (neutral, monoisotopical)	Estimated elemental formula <sup>b</sup>	RT <sup>c</sup> (min)	Tentative identity (name of possible compounds) <sup>d</sup>	Peak area <sup>e</sup>
37	372.2295	C23 H32 O4	10.11	(+/-)-4-Acetoxycannabichromene/acetyl cannabigeroinol/7.8-dehydro-10-O-ethylcannabitriol/	9.7E+04
38	384.3021	C26 H40 O2	10.29	Sesquicannabigerol/O-propylcannabidiol/O-pentyl-delta9-trans-tetrahydrocannabinol	7.3E+05
39	326.2237	C22 H30 O2	10.61	Confluentin	4.1E+05
40	384.3019	C26 H40 O2	10.81	Sesquicannabigerol/O-propylcannabidiol/O-pentyl-delta9-trans-tetrahydrocannabinol	2.2E+05
41	328.2390	C22 H32 O2	10.97	O-Methylcannabidiol	3.7E+04
42	640.4486	C43 H60 O4	12.57	Cannabisol	7.5E+05
43	494.3386	C32 H46 O4	13.35	beta-Fenchyl delta 9 - tetrahydrocannabinolate/epi-bornyl delta 9 - tetrahydrocannabinolate/alpha - terpenyl delta 9 - tetrahydrocannabinolate/4-terpenyl delta 9 - tetrahydrocannabinolate/bornyl delta 9 - tetrahydrocannabinolate/alpha - fenchyl delta 9 - tetrahydrocannabinolate	2.0E+05

<sup>a</sup> Mass error compared the theoretical exact mass < 5 ppm

<sup>b</sup> Match of isotopic pattern confirmed

<sup>c</sup> Retention time

<sup>d</sup> Tentative identification of compounds based on available scientific articles (Hanus̄ et al. 2016, Mechoulam 2002); possible identities are separated by the symbol “/”

<sup>e</sup> Peak area related to the undiluted sample prepared according to stated procedure

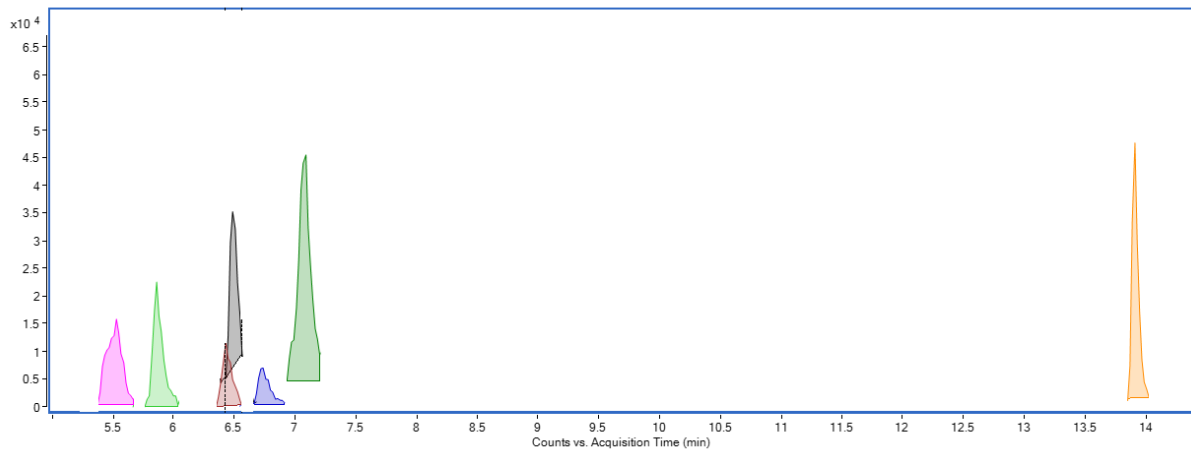
### B) Targeted screening of other (non-cannabinoid) biologically active compounds

In the tested samples, the following amounts of other (non-cannabinoid) biologically active compounds (**Table VI**) characterized by unique combination of exact mass and retention time RT were detected.

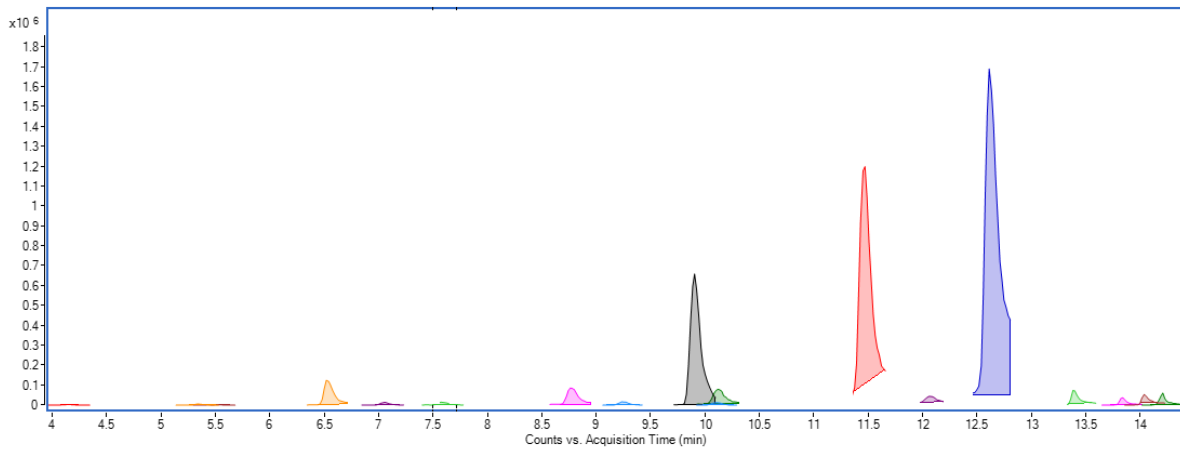
**Table VI:** Number of other biologically active compounds detected in the sample in ESI+ and ESI-

Sample name	Number of other biologically active compounds detected in ESI+	Number of other biologically active compounds detected in ESI-
ML 4912/21	7	18

Extracted ion chromatograms (XIC) of the detected compounds in both ionization modes (ESI+, ESI-) are documented on **Figures 7 - 8**. All detected non-cannabinoid biologically active compounds (tentative identification) are also summarized in **Table VII** (ESI+) and **Table VIII** (ESI-). Considering the possible existence of isomeric forms, one detected compound may have several identities.



**Figure 7:** ML 4912/21 - UHPLC-HRMS/MS extracted ion chromatogram of 7 compound with unique combination of exact mass and RT detected in ESI+ (obtained for undiluted sample)



**Figure 8:** ML 4912/21 - UHPLC-HRMS/MS extracted ion chromatogram of 18 compounds with unique combination of exact mass and RT detected in ESI- (obtained for undiluted sample)

**Table VII:** Overview of compounds detected in ESI+ (corresponding with the Figure 7)

Cmp. number	Measured exact mass <sup>a</sup> (neutral, monoisotopical)	Estimated elemental formula <sup>b</sup>	RT <sup>c</sup> (min)	Tentative identity (name of possible compounds) <sup>d</sup>	Compound group	Peak area <sup>e</sup>
1	302.1158	C17 H18 O5	5.49	4.5-dihydroxy-2.5-dihydroxy-2.3.6-trimethoxy-9.10-dihydrophenanthrene	new non-cannabinoid constituent f	1.5E+05
2	272.1054	C16 H16 O4	5.86	cannithrene-2	phenol	1.3E+05
3	368.1266	C21 H20 O6	6.42	cannflavin / isocannflavin B	flavonoid	8.1E+04
4	204.1878	C15 H24	6.49	beta-caryophyllene / trans-gamma-bisabolene / cis-gamma-bisabolene / cis-beta-farnesene / allo-aromadendrene / viridiflorene / trans-alpha-farnesene / alpha-humulene (alpha-caryophyllene) / alpha-guaiene / beta-selinene / alpha-selinene / gamma-murolene / gamma-curcumene / alpha-ylangene / beta-elemene / alpha-cis-bergamotene / alpha-trans-bergamotene / alpha-cadinene / alpha-longipinene / alpha-copaene	terpenoid	2.1E+05
5	222.1985	C15 H26 O	6.73	guajol / trans-nerolidol / gamma-eudesmol / beta-eudesmol / alpha-eudesmol / epi-alpha-bisabolol	terpenoid	4.7E+04
6	312.1740	C20 H24 O3	7.06	cannabistilbene-I	phenol	2.3E+05
7	412.3701	C29 H48 O	13.91	stigmasterol	phytosterol	1.7E+05

<sup>a</sup> Mass error compared the theoretical exact mass < 5 ppm

<sup>b</sup> Match of isotopic pattern confirmed

<sup>c</sup> Retention time

<sup>d</sup> Possible identities are separated by the symbol “/”

<sup>e</sup> Peak area related to the undiluted sample prepared according to stated procedure

<sup>f</sup> Mohamed M. Radwan et al. 2008

**Table VIII:** Overview of compounds detected in ESI- (corresponding with the Figure 8)

Cmp. number	Measured exact mass <sup>a</sup> (neutral, monoisotopical)	Estimated elemental formula <sup>b</sup>	RT <sup>c</sup> (min)	Tentative identity (name of possible compounds) <sup>d</sup>	Compound group	Peak area <sup>e</sup>
1	188.1048	C9 H16 O4	4.15	azealic acid	fatty acid	9.0E+04
2	246.1255	C15 H18 O3	5.33	cannabispiran / isocannabispiran	phenol	3.6E+04
3	242.0940	C15 H14 O3	5.48	isocannabispiradienone / cannithrene-1	phenol	2.1E+04
4	144.1151	C8 H16 O2	6.52	caprylic acid	fatty acid	8.3E+05
5	158.1307	C9 H18 O2	7.04	pelargonic acid	fatty acid	5.1E+05
6	172.1463	C10 H20 O2	7.58	capric acid	fatty acid	1.3E+05
7	200.1777	C12 H24 O2	8.76	lauric acid	fatty acid	2.8E+05
8	276.2084	C18 H28 O2	9.24	stearidonic acid	fatty acid	1.1E+05
9	278.2248	C18 H30 O2	9.90	alpha-linolenic acid / gamma-linolenic acid / isolinolenic acid	fatty acid	4.6E+06
10	372.2294	C23 H32 O4	10.10	5-acetoxy-6-geranyl-3-n-pentyl-1.4-benzoquinone	new non-cannabinoid constituent <sup>f</sup>	9.7E+04
11	228.2089	C14 H28 O2	10.11	myristic acid	fatty acid	9.4E+04
12	256.2405	C16 H32 O2	11.46	palmitic acid	fatty acid	6.9E+07
13	270.2557	C17 H34 O2	12.06	margaric acid	fatty acid	2.4E+04
14	284.2719	C18 H36 O2	12.61	stearic acid	fatty acid	1.1E+08
15	312.3024	C20 H40 O2	13.39	arachidic acid / isoarachidic acid	fatty acid	1.8E+05
16	340.3335	C22 H44 O2	13.83	behenic acid	fatty acid	6.3E+05
17	414.3849	C29 H50 O	14.04	beta-sitosterol	phytosterol	3.6E+04
18	368.3647	C24 H48 O2	14.20	lignoceric acid	fatty acid	1.2E+05

<sup>a</sup> Mass error compared the theoretical exact mass < 5 ppm

<sup>b</sup> Match of isotopic pattern confirmed

<sup>c</sup> Retention time

<sup>d</sup> Possible identities are separated by the symbol "/"

<sup>e</sup> Peak area related to the undiluted sample prepared according to stated procedure

<sup>f</sup> Mohamed M. Radwan et al. 2008

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*End of the appendix*